### JSS MAHAVIDYAPEETA JSS COLLEGE OF ARTS, COMMERCE AND SCIENCE

(An autonomous College of University of Mysore, Re-accredited by NAAC with 'A' grade) B.N. Road, Mysuru-570 025, Karnataka



# CAREER ORIENTED COURSE IN INDUSTRIAL MICROBIOLOGY

#### INDUSTRIAL MICROBIOLOGY SYLLABUS

(W.E.F. 2022-2023)
Proposed regulations for the Career Oriented Course in Industrial Microbiology

- **Rule 1 :** A candidate for admission to the career oriented (Add on course) must have passed PUC (10+2) or equivalent examination with science subjects. The rules/regulations for admission for the course shall be in accordance with those framed by the college from time to time.
- Rule 2: The course of study for the career oriented course is an add on course. Any graduate studying student with science subjects may be admitted.
- **Rule 3 :** The course of study is taught in three years under semester scheme. The first year course is entitled "CERTIFICATE COURSE IN INDUSTRIAL MICROBIOLOGY". It shall be of one academic year with two semesters / terms of 16 weeks each.

The second year course is entitled "DIPLOMA COURSE IN INDUSTRIAL MICROBIOLOGY". It shall be of one academic year with two semesters / terms of 16 weeks each. However, to join this course the candidate should have completed the Certificate Course.

The third year course is entitled "ADVANCED DIPLOMA IN INDUSTRIAL MICROBIOLOGY". It shall be of one academic year with two semesters/terms of 16 weeks each. However, to join this course the candidate should have completed the Diploma Course.

- **Rule 4 :** In the sixth semester the candidate should undertake a project work / Industrial training.
- **Rule 5 :** A candidate is awarded with the respective certificates at the end of each course with the marks obtained.
- **Rule 6 :** A candidate will be permitted to appear for the examination provided he / she has attended both the semesters for each said course and has a minimum attendance of 75%, as per rules.
- Rule -7: For the purpose of awarding the certificate the theory component and

practical component is 70 + 30 marks respectively. In each semester under theory paper 50 marks for the actual examination and 20 marks for the Internal assessment. For the purpose of IA for theory, the college will conduct at least one test in each term. Out of 30 marks in the practical component 20 marks for the actual practical examination 05 marks for record and 05 marks for I.A. Internal Assessment for practicals should be based on performance of the experiments in each class. The experiments conducted by the candidate and results obtained should be taken into consideration. At the end of semester the concerned teacher shall take the average of marks awarded in each practical and submit it to the HOD / Course Co-ordinator.

**Rule -8:** The VI Semester shall comprise exclusively a project work / industrial training to be performed by the candidate and it will be evaluated at the end for 100 marks.

#### ${\bf J.S.S.~COLLEGE~OF~ARTS,~COMMERCE~AND~SCIENCE,~OOTY~ROAD,~MYSORE-25}$

(Autonomous)

Scheme of Study and Examination for the Career Oriented Course in Industrial Microbiology (Add on course )

FIRST YEAR: Certificate Course in Industrial Microbiology

	This Think Colombate Southern Industrial Whet obliding								
SEM	Paper	Title of the paper	Teaching w	Marks for examination Maximum 100 marks					
Scheme			Theory	Practical	Theory max. 70		Practical max. 30		
					Ex.	IA	Ex.	IA	REC
I Paper code	1 IMA010	Tools and techniques in Industrial Microbiology	02	02	50	20	20	05	05
II Paper code	2 IMB010	Microbial Physiology and Genetics	02	02	50	20	20	05	05

**SECOND YEAR: Diploma Course in Industrial Microbiology** 

SECOTO TENENT CONTROL IN THE CONTROL IN THE CONTROL OF THE CONTROL									
SEM Paper		Title of the paper	Teaching hours per weeks		Marks for examination Maximum 100 marks				
			***	CCKS			1111 100	) marks	
Scheme			Theory	Practical	Theory max. 70		Practical max. 30		
				Ex.	IA	Ex.	IA	REC	
III	3	Fermentation	02	02	50	20	20	05	05
Paper	IMC010	technology							
code									
IV	4	Environmental	02	02	50	20	20	05	05
Paper	IMD010	Microbiology							
code									

THIRD YEAR: Advanced Diploma Course in Industrial Microbiology

	THEO TEAK. Advanced Diploma Course in industrial witchblology								
SEM Paper		Title of the paper	Teaching hours per		Marks for examination				
SEM	rapei	Title of the paper	weeks		Maximum 100 marks				
Scheme			Theory	Practical	Theory max. 70		Practical max. 30		
					Ex.	IA	Ex.	IA	REC
V	5	Gene Technology	02	02	50	20	20	05	05
Paper	IME010	and Patents							
code									
VI	6	Industrial Training /	NIL	NIL	Max.Marks 100		Report: 60		
Paper	IMF010	Project					20 : Presentation		ntation
code							20: viva-voce		

Total: 200 + 200 + 200 = 600 Marks

### CERTIFICATE COURSE IN INDUSTRIAL MICROBIOLOGY I SEMESTER

#### PAPER - I: TOOLS AND TECHNIQUES IN INDUSTRIAL MICROBIOLOGY

Paper code: IMA010 THEORY 28 Hours

#### **COURSE OUTCOME:**

After successful completion of the course students are able to:

- **CO1.** Gain basic knowledge about Industrial Microbiology starting from history and learn the over view of scope and importance of industrially important microbes.
- CO2. Learn about the taxonomical classification of Microbes.
- CO3. Demonstrate theory and practical skills in microscopy, their handling techniques.
- **CO4.** Gain skills of isolation, culturing, strain improvement and maintenance of pure culture.

Unit –I 7Hours

- General introduction, Scope of industrial microbiology. History and development of industrial microbiology. Experiments of Louis Pasteur, Robert Koch and Alexander Fleming.
- 2. Isolation, identification and characteristics of microbes of industrial importance. Classification of bacteria, fungi, actinomycetes and viruses.

Unit –II 7Hours

- 3. Microscopy Construction and working principles of bright field, phase contrast, Fluorescent microscope. A brief account on Electron Microscopy
- 4. Principles of sterilization: Sterilization equipments, sterilization of media, air and other ancillary equipments.

Unit –III 7Hours

- 5. Substrates for industrial fermentation- Raw materials, Energy sources, Mineral sources, Antifoam agents and Media formulations.
- 6. Batch and Continuous Fermentations, Surface, Submerged and Solid-substrate fermentations, Scale-up of Fermentation.

Unit –IV 7Hours

- **7. Strain development :** Isolation, screening techniques, improvement of strains by mutation; recombination, gene regulation, technology and other genetic methods.
- **8.** Preservation and maintenance of industrial microorganisms.

**Total Marks 70: 60(Theory + 10 (Internal Assessment-Test)** 

**I SEMESTER** 

#### PRACTICAL: I

### PAPER - I: TOOLS AND TECHNIQUES IN INDUSTRIAL MICROBIOLOGY 10 Practicals(1 Practical of 2 hours per week)

- 1. Laboratory safety: Good laboratory practices.
- 2. Study of simple and compound microscopes, their handling, including oil immersion objective.
- Demonstration of laboratory equipments: Pressure cooker, Autoclave, Hot-air oven, Incubator, Inoculation chamber / Inoculation hood, Inoculation loop, Inoculation needle, Membrane filter and Colony counter.
- 4. Demonstration of pH meter, Colorimeter, Absorption and emission spectroscopy and Turbidometer.
- 5. Preparation of media: Nutrient agar and Potato Dextrose Agar.
- 6. Cultivation of microorganisms on agar plate (point inoculation), broth, anaerobic cultivation (candle jar / gas pack method).
- 7. Isolation of microorganisms by spread plate, pour plate and streak plate methods.
- 8. Maintenance of stock culture on fresh agar slants; overlaying with mineral oil and stab culture.
- 9. Staining and mounting of algae (*Chlorella*) and fungi (*Pencillium*).
- 10. Staining of bacteria (simple and differential staining).

Total Marks 30:20 (Practicals)+05 (Internal Assessment-Test)+05 (Record)

#### II SEMESTER

### PAPER - II: MICROBIAL PHYSIOLOGY AND GENETICS Paper code: IMB010 THEORY 28hours

#### **COURSE OUTCOME:**

After successful completion of the course students are able to:

- **CO1.** Understand the basic microbial structure, function and study of the comparative characteristics of prokaryotes and eukaryotes.
- **CO2.** Inculcate the knowledge regarding microbial growth, functions, physiology and metabolism
- CO3. Get equipped with various methods of bacterial growth measurement
- **CO4.** Understand the experimental evidences to prove DNA as genetic material and differentiate various method of recombination in bacteria

Unit-I 7Hours

- 1. **Microbial cell structure and function**-prokaryotes, eubacteria and fungi with reference to their cell membrane, cell wall and internal cellular organization.
- 2. **Microbial nutrition**: Nutritional requirements of microorganisms uptake of nutrients- active transport, passive transport, facilitate diffusion, group translocation and iron uptake.

Unit-II 7Hours

- 3. Microbial growth and generation time, growth curve-phases of growth and their significance. Measurement of growth by cell number and biomass; direct microscopic count (Haemocytometer, turbidometer and spectrophotometer method).
- 4. **Microbial Metabolism:** Photosynthetic microbes, oxygenic and non-oxygenic photosynthesis, photophosphorylation, Calvin cycle dark reaction. Respiratory pathways Glycolysis, Kreb's cycle, pentose phosphate pathway, oxidative and substrate level phosphorylation.

Unit-III 7Hours

5. Chromosomes-prokaryotic and Eukaryotic organization. Cell division-mitosis, meiosis and cell cycle regulation. Recombination in bacteria; Transformation, Transduction (types) and Conjugation process, F-factor, Transposons, Extrachromosomal genetic elements and their importance.

Unit-IV 7Hours

6. Chemical basis of heredity-evidence for DNA and RNA as genetic material. Watson and Crick model of DNA, structure and its functional importance. DNA replication- mode and mechanism of Replication in prokaryotes and eukaryotes.

Total Marks 70:20(Theory)+10(Internal Assessment-Test)

#### II SEMESTER PRACTICAL:II

### PAPER - II : MICROBIAL PHYSIOLOGY AND GENETICS 10 Practicals(1 Practical of 2 hours per week)

- Acid and gas production from Carbohydrates demonstration of fermentation of Lactose.
- 2. Starch hydrolysis.
- 3-5. Qualitative tests for the detection of macromolecules.
  - Glucose Benedict's test, Molisch's test, Fehlings test, Picric acid test.
  - Protein Biurette test, Ninhydrin test, Millon's test, Xanthoproteic test.
  - Lipids Acrolein test, Sudan III test, Emulsification test and Solubility test.
- 6. Effect of temperature on growth of microorganisms.
- 7. Effect of pH on growth of microorganisms.
- 8. Measurement of growth by cell number using Haemocytometer and cell mass by turbidometer/ spectrophotometer method.
- 9. Isolation of DNA.
- 10. Demonstration of the following models / photographs of DNA, Transcription and Translation, DNA replication, t-RNA.

Total Marks 30: 20(Practicals)+05 (Internal Assesment-Test)+05(Record)

### DIPLOMA COURSE IN INDUSTRIAL MICROBIOLOGY III SEMESTER

**PAPER-III: FERMENTATION TECHNOLOGY** 

Paper code: IMC010 THEORY 28hours COURSE OUTCOME:

After successful completion of the course students are able to:

- **CO1.** Acquaint with different types of fermentation processes and equipment and underlying principles in downstream processing
- CO2. Understand how microbiology is applied in manufacture of industrial products
- **CO3.** Understand the beneficial role of microorganisms in fermented dairy products & in alcoholic beverages
- **CO4.** Gain knowledge about biofertilizers and biopesticide in agriculture and understand various plant microbes interactions and their applications

Unit-I 7Hours

- 1. **Introduction to Fermentations**: Design of typical fermenters, devices for aeration and agitation, Seed and Pilot Scale fermenters.
- Down Stream Processing: Recovery of fermentation products-precipitation, filtration, centrifugation, distillation, cell disruption, solvent recovery, drying and crystallization.

Unit-II 7Hours

3. Industrial production and their uses :

Enzymes: Amylase

Vitamins - Vitamin B-12

Antibiotics: Penicillin.

**SCP**: Spirullina

Organic acids: Citric acid.

Production of fuels: Ethanol

**Health care products:** Steroid transformation, Monoclonal antibodies.

Unit-III 7Hours

4. **Food fermentations**: Alcoholic beverages, Cheese making, Bread making, Fermented soya based foods. Meat fermentation, Fermented milk and mushroom cultivation.

Unit-III 7Hours

5. **Microbes-Plant interaction-** Mycorrhizae-Biology and applications, **Biofertilizers**, microbial inoculants, Brief account of production and application of *Rhizobium*, *Azotobacter*, *phosphobacteria* and *Cyanobacteria*.

**Biopesticides:** Fungal, Bacterial and Viral Biopesticides and their applications.

#### Total Marks 70:60(Theory)+10(Internal Assesment-Test

## III SEMESTER PRACTICAL: III PAPER-III: FERMENTATION TECHNOLOGY 10 Practicals (1 Practical of 2 hours per week)

- 1. Isolation and identification of microorganisms of spoiled fruits (fungi and bacteria).
- 2. Isolation of Aspergillus flavus from spoiled food.
- 3. MBRT test for microbial contamination of milk.
- 4. Isolation of *Lactobacilli* and *Streptococcus* from curds.
- 5. Production of citric acid from Aspergillus niger.
- 6. Production of wine from grapes.
- 7. Production of alcohol from jaggery / Molasses.
- 8. Estimation of Percentage alcohol in a given sample by specific gravity bottle method.
- 9. Mushroom cultivation.
- 10. Antibiotic sensitivity test.

Total Marks 30:20(Practicals)+05 (Internal Assesment-Test)+05(Record)

#### **IV SEMESTER**

#### PAPER - IV: ENVIRONMENTAL MICROBIOLOGY

Paper code: IMD010 THEORY 28hours

#### **COURSE OUTCOME:**

After successful completion of the course students are able to:

- CO1. Know the role of microorganisms in soil, air, water, waste water and bioremediation
- **CO2.** Learn the occurrence, abundance and distribution of microorganisms in the environment and their role in the environment
- **CO3.** Know about bioaerosols, airsample collection and analysis and **co**ntrol measures of air borne microbes
- **CO4.** Understand the basic principles of bioremediation

Unit-I 7Hours

 Marine and freshwater microorganisms. The microbial community in marine and freshwater environments. Water purification, Microbiological analysis of water. Waste water and effluent treatment - treatment methods – Primary, Secondary and tertiary treatment. Sludge treatment and disposal, Composting and Ensiling.

Unit-II 7Hours

- 2. **Nutrient cycling processes:** Biogeochemical cycles: Nitrogen cycle, Phosphorous cycle and Carbon cycle.
- 3. **The environments of soil microorganisms:** Associated soil microorganisms with plants. Microbial interaction: Neutralism, Mutualism, Parasitism, Antagonism, Comensalism, Predation, Synergism, and competition. Rhizosphere, Actinorrhizae and Mycorrhizae.

Unit-III 7Hours

- 4. **Aerobiology**: Air-borne microbes, Sampling techniques-Andersen sampler, Rotorod sampler, Hirst spore trap and Vertical cylinder spore trap. Aerobiology and Human health.
- 5. **Microbes and degradation of wastes:** Treatment of solid and liquid industrial wastes, Microbial degradation of pesticides and Xenobiotics, Bioremediation.

Unit-IV 7Hours

- 6. **GeoMicrobiology:** Microbes in metal extraction, Mineral leaching and mining.
- 7. **Production of biopesticides** (microbial insecticides)
  - Bacterial, Viral, Fungal and protozoa.
  - Production of microbial herbicides.

Total Marks 70: 60(Theory)+10(Internal Assessment-Test)

### IV SEMESTER PRACTICAL:IV

#### PAPER - IV: ENVIRONMENTAL MICROBIOLOGY

10 Practicals(1 Practical of 2 hours per week)

- 1. Microscopic observations of different water samples for biological indicator of water pollution.
- 2. a) Standard analysis of water.
  - b) Determination of MPN.
- 3. IMViC tests.
- 4. Viable count SPC and Membrane Filter Technique.
- 5. Determination of BOD
- 6. Estimation of total solids in sewage.
- 7. Measurement Cl<sub>2</sub>, PO<sub>4</sub> and NO<sub>3</sub> in water.
- 8. Demostration of Production of Vermicompost.
- 9. Isolation of free-living N<sub>2</sub> fixers from soil.
- 10. Visit to Effluent treatment plant.

**Total Marks 30: 20 (Practicals) + 05 (Internal Assessment-Test) + 05(Record)** 

### ADVANCED DIPLOMA COURSE IN INDUSTRIAL MICROBIOLOGY V SEMESTER

#### PAPER - V: GENE TECHNOLOGY AND PATENTS

Paper code: IME010 THEORY 28Hours

#### **COURSE OUTCOME:**

After successful completion of the course students are able to:

- **CO1.** Know genetics of microorganisms and recombinant DNA technology used in microbiological research
- CO2. Understand the properties, structure and function of genes in microorganisms
- **CO3.** Understand techniques, social and ethical issues concerning genetic engineering. Applications of genetic engineering in various fields.
- CO4. Gain knowledge about Patent.

Unit-I 7 hours

- 1. **Genetic Engineering:** Definition, concept and scope of genetic engineering; Milestones in Recombinant DNA Technology. Gene technology and industrial microbiology.
- 2. **Recombinant DNA Technology:** Restriction enzymes; Isolating and cloning fragments; Isolating and purifying cloned DNA, Genomic and C-DNA Libraries and Cloning vectors.

Unit-II 7 hours

- 3. **Selection of recombinants:** Expression of foreign genes in cloned organisms.
- 4. **Analysis of gene products :** (a) Nucleic acid hybridization, (b) Sequencing of proteins, and nucleic acids, Southern, Northern and Western blotting techniques, PCR and DNA finger printing.

Unit-III 7 hours

- 5. **Genetic engineering :** Safety, Social, Legal and Ethical considerations. Risk in genetic engineering. Microbial transformation: Types of bioconversion reactions, procedure for biotransformation and transformation of antibiotics.
- 6. Applications of genetic engineering:
  - a) Medical,
  - b) Industrial,
  - c) Environmental and
  - d) Agricultural Applications in brief

Unit-IV 7 hours

7. **Patents;** patent regulations of processes, products and microorganisms, Entrepreneurship and marketing of products.

**Total Marks 70: 60(Theory) + 10(Internal Assessment-Test)** 

### V SEMESTER PRACTICAL: V

### PAPER - V: GENE TECHNOLOGY AND PATENTS 10 Practicals(1 Practical of 2 hours per week)

- 1. Isolation of genomic DNA and its quantification.
- 2. Isolation of RNA and its quantification.
- 3. Salt fractionation of yeast proteins and quantification.
- 4. Separation of proteins by SDS-PAGE.
- 5. Isolation of Streptomycin resistant mutant of *E. coli*.
- 6. Isolation of plasmid from bacteria purification by agarose gel electrophoresis.
- 7. Preparation of competent *E. coli* cells.
- 8. Transformation and recovery of plasmid clones.
- 9. Demonstration of Southern blotting and Northern blotting.
- 10. Demonstration of PCR Technology.

**Total Marks 30: 20(Practicals) + 05 (Internal Assessment-Test) + 05(Record)** 

#### Paper code: IMF010 VI SEMESTER

#### PAPER-VI: INDUSTRIAL TRAINING / PROJECT WORK

The candidate need to work for a period of 16 weeks in any recognized Industry / University, under the guidance of an expert in the field of Industrial Microbiology. At the end of the semester candidate need to submit a report duly certified by the authority of the industry / Academic institution. The same should be certified by the HOD / Co-ordinator. The project work will be evaluated for 100 marks (60 for report and 20 for presentation and 20 for Viva-Voce examination)

Signature of the Co-Ordinator Microbiology

### SCHEME OF THEORY EXAMINATION FOR UGC CAREER ORIENTED COURSE IN INDUSTRIAL MICROBIOLOGY

(Applicable to Semester I to V)

	Time: 03 Hrs	Max. Marks: 50
1	Answer any TEN of the following: (12 questions to be given and 10 to be answered)	1 x 10 =10
	a)	
	b)	
	c)	
	d)	
	e)	
	f)	
	g)	
	h)	
	i)	
	j)	
	k)	
	1)	
2.	Answer any FOUR of the following	$4 \times 4 = 16$
	(6 questions to be given and 4 to be answered) – short answer type	
	a)	
	b)	
	c)	
	d)	
	e)	
	f)	
3.	(3, 4, 5 essay type questions – Each with internal choices)	$8x\ 3 = 24$

### SCHEME OF PRACTICAL EXAMINATION FOR CAREER ORIENTED COURSE IN INDUSTRIAL MICROBIOLOGY

#### (Applicable to Semester I, II, III, IV and V)

Time: 03 Hrs
 Max. marks: 25(20 + 05)
 Conduct experiment 'A' assigned, tabulated data, analyse and draw conclusions
 Write a critical notes on B, C and D. (3 x 2) = 06
 Identify E, F and G with reasons. (3 x 2) = 06
 Class Record 05

Internal assessment-05

#### **REFERENCES**

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List of Examiners Recommended by the BOS in Microbiology

Sl No.	Name	mmended by the BOS in Microbiology  College address
1	DrM .Seema	Chairperson, Dept. of Microbiology
		JSS College, Ooty road,
		Mysore
2	H.P.Spoorthy	Assistant prof.
		Dept. of Microbiology
		JSS College,
		Ooty road,
		Mysore
3	Dr.S.Mahadevamurthy	Associate Prof & HOD
		Dept. of Microbiology
		Yuvaraja's college
		Mysore.
4	Dr.Syeda Kauser Fathima	Associate Prof. of Microbiology
		Maharani's Science College for women
		JLB road
		Mysore.
5	Dr. H.S. Jayanth.	Asso.Prof.of Microbiology
		Dept. of Microbiology
		Yuvaraja's college
		Mysore.
6	Dr.Uma Maheshwari	Assistant prof.
		JSS University,
		JSSAHER, Mysuru
7	Sri. M. Girish	Assistant prof.
		Dept. of Microbiology
		JSS College for Women
		Saraswathipuram, Mysore
8	Dr. P.K.Maheshwar	Assistant Prof.
		Dept. of Microbiology
		Yuvaraja's college, Mysore.
9	Smt. M.S.Shobha	Assistant Prof,
		Dept. of Microbiology
		Maharani's Science College
		Mysore
10	Sri. R.A. Manjunath	Assistant Prof.
	j	Dept. of Microbiology
		Saradavilas College, Mysore
11	Dr.M.P. Ragavendra	Assistant Prof.
		Dept. of Microbiology
		Maharani's Science College, Mysore

12	Dr.K.Girish	Assistant Prof.
		Dept. of Microbiology
		Maharani's Science College,
		Mysore
13	Sri. G.S. Siddegowda	Assistant Prof.
		Dept. of Microbiology
		Maharani's Science College
		Mysore
14	Dr.N.S.Devaki	Assistant Prof.
		Dept. of Molecular Biology
		Yuvaraja's College, Mysore
15	Syeda Farahna Parveen	Assistant Prof.
		Dept. of Microbiology
		St.Philomina's College, Mysore
16	Smt. Vanitha	Assistant Prof.
		Dept. of Microbiology
		Maharani's Science College, Mysore
17	Smt. Revanamba	Assistant Prof.
		Dept. of Microbiology
		Maharani's Science College, Mysore
18	Mahadevaprasad	Assistant prof.
		Dept. of Microbiology
		JSS College for Women
		Saraswathipuram, Mysore
19	Dr.Nagalambika	Assistant prof.
		JSS University,
		JSSAHER, Mysuru
20	RajaRajeshwari.R	Assistant prof.
		Dept. of Microbiology
		SDM College
		JLB Road,Mysuru
21	Uzma Bathool	Assistant Prof.
		Dept. of Microbiology
		St.Philomina's College, Mysore
22	Shruthi Prakash	Assistant Prof.
		Dept. of Microbiology
22		Mahajana's Science College, Mysuru
23	Athiya sultan	Assistant prof.
		Dept. of Microbiology
		SDM College
		JLB Road, Mysuru
24	C.Poornima Devi	Assistant Prof.
		Dept. of Microbiology
		Yuvaraja's college,Mysore

	Dr.Rakshith	Assistant Prof.
25		Dept. of Microbiology
		Yuvaraja's college, Mysore
	Assistant Prof.	
26		Dept. of Microbiology
		Maharani's Science College, Mysore
	Niveditha Prakash	Assistant prof.
27		Dept. of Microbiology
JSS Co		JSS College for Women
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	Dr.K.Sumana	Assistant prof.
28		JSS University, JSSAHER ,Mysuru
Samjna.S.R Assistant Prof.		Assistant Prof.
29 Dept. of Microbiology		Dept. of Microbiology
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#### DEPARTMENT OF MICROBIOLOGY

#### LIST OF BOS MEMBERS (2020-23)

	MEMBERS	SIGNATURE
	Dr. M. Seema-Chairperson	
1	Assistant Professor and Head	
	Department of Microbiology	
	JSS College, Ooty road, Mysuru-25	
	Dr. Shubha Gopal	
2	Professor	
	DOS in Microbiology	
	Manasagangotri, Mysuru	
	Member (VC Nominee)	
	Dr.C. R. Rekha	
3	Associate Professor	
	DOS in Microbiology	
	Government Science First Grade College	
	Yelahanka	
	Bengaluru	
	Member (VC Nominee)	
	Dr. Syeda Kousar Fathima	
4	Associate Professor	
	PG Department of Microbiology	
	Maharani's Science College, Mysuru	
	Member (VC Nominee)	
	Dr. Srilakshmi Desiraju	
5	TRIPHASE Pharmaceuticals pvt. Ltd	
	KSSIDC, Hebbal Industrial area, Mysuru	
6	Chethana. G	
	Jnanasarovara International Residential School	
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