JSS COLLEGE OF ARTS, COMMERCE AND SCIENCE (AUTONOMOUS) OOTY ROAD, MYSORE – 25.



CURRICULUM FOR

For B.Sc., (Basic/ Hons.) Degree

BIOTECHNOLOGY

(As per NEP-2020 Model Curriculum)

Implementation Year 2021-22

B.Sc., (Basic/ Hons.) Degree

Scheme of study for B.Sc. Biotechnology As per NEP-2020 Model from 2021-22

YEAR	SEMESTER	CORE COURSE	COURSE CODE	TITLE OF THE PAPER	NO. OF CREDITS	LECTURE/ PRACTICAL/ HOUR/WEEK	TOTAL TEACHING HOURS
	Ţ	DSC -I:Theory		Cell biology & genetics	4	4	56
	1	DSC -I:Pract		Cell biology & genetics	2	4	56
I BSc		OE:Theory		Biotechnology for human welfare	3	3	42
1 DSC		SEC-: Theory		Biotechnological Skills and Analytical Techniques	1	1	14
	II	DSC-II:Theory		Microbiological Methods and Techniques	4	4	56
	11	DSC-II: Pract		Microbiological Methods and Techniques	2	4	56
		OE:Theory		Applications of Biotechnology in Agriculture	3	3	42

JSS COLLEGE OF ARTS, COMMERCE AND SCIENCE, OOTY ROAD, MYSORE Scheme of Examination for B.Sc. Biotechnology As per NEP-2020 Model from 2021-22

Year	Semester	Core course	ore course Course code Title of the paper		credits	Maximum Marks in exam/Assessment					Exam Duratio n	
					L:T:P	I		Total	Th	Pr		
						C-1	C-2	C-3				
		DSC -		Cell biology & genetics	4: 0: 0							
I B.Sc		I:Theory				20	20	60	100	3h	3h	
I B.SC	I	DSC -I:Pract		Cell biology & genetics	0: 0: 2	10	15	25	50			
		OE:Theory		Biotechnology for human welfare	3:0:0							
						20	20	60	100	3h	3h	
		SEC-: Theory		Biotechnological Skills and Analytical Techniques	0:0:1							
		DSC-		Microbiological Methods and Techniques	4: 0: 0							
	п	II:Theory				20	20	60	100	3h	3h	
		DSC-II: Pract		Microbiological Methods and Techniques	0: 0: 2	10	15	25	50			
		OE:Theory		Applications of Biotechnology in Agriculture	3:0:0							
						20	20	60	100	3h	3h	

JSS COLLEGE OF ARTS, COMMERCE AND SCIENCE, OOTY ROAD, MYSORE Scheme of Examination Programme – B.Sc., BMBt; Programme code –BSC

Year	Semester	Semester Core course	Core course Course code Title of the paper	Title of the paper	credits	N	Exam Duration				
					L:T:P	IA(Th	• • • • • • • • • • • • • • • • • • • •		Total	Th	Pr
						C-1	C-2	C-3			
	I	DSC-I :Theory	DMA22006	Cell Biology and Genetics	4: 0: 0	15	15	70	100	3h	3h
I B.Sc		DSC-I: Pract	DMA22106	Cell Biology and Genetics	0: 0: 2	7.5	7.5	35	50		
	II	DSC-II:Theory	DMB22006	Biomolecules and Bio-analytical techniques	4:0:0	15	15	70	100	3h	3h
		DSC-II: Pract	DMB22106	Biomolecules and Bio-analytical techniques	0:0:2	7.5	7.5	35	50		
	ш	DSC-III:Theory	DMC22006	Molecular biology and Genetic engineering	4:0:0	15	15	70	100	3h	3h
II B.Sc		DSC-III: Pract	DMC22106	Molecular biology and Genetic engineering	0:0:2	7.5	7.5	35	50		
	IV	DSC-IV: Theory	DMD22006	Plant Tissue culture and Animal Cell culture		15	15	70	100	3h	3h
		DSC-IV: Pract	DMD22106	Plant Tissue culture and Animal Cell culture	0:0:2	7.5	7.5	35	50		
	v	DSE: Elective 1:Theory	DME22006	Elective 1: Immunology and Medical Biotechnology	4:0:0	15	15	70	100		
	'	Elective 1: Pract	DME22106	Elective 1: Immunology and Medical Biotechnology	0:0:2	7.5	7.5	35	50	3h	3h
		Elective 2: Theory	DME22206	Elective 2: Microbial technology and Agricultural Biotechnology	4:0:0	15	15	70	100		
III B.Sc.		Elective 2: Pract	DME22306	Elective 2: Microbial technology and Agricultural Biotechnology	0:0:2	7.5	7.5	35	50		
		SEC 1: Theory	DME22406	SEC 1:Microbial Techniques	2:0:0	7.5	7.5	35	50	2h	-
		SEC 2: Theory	DME22506	SEC 2:Enzymology	2:0:0	7.5	7.5	35	50	2h	-
	VI	DSE: Elective 1:Theory	DMF22006	Elective 1: Environmental Biotechnology and Biostatistics	4:0:0	15	15	70	100		
		Elective 1: Pract	DMF22106	Elective 1: Environmental Biotechnology and Biostatistics	4:0:0	15	15	70	100	3h	3h
		Elective 2: Theory	DMF22206	Elective 2: Bioinformatics and Bioprocess technology	4:0:0	15	15	70	100	-	
		Elective 2: Pract	DMF22306	Elective 2: Bioinformatics and Bioprocess technology	4:0:0	15	15	70	100		

MODEL CURRICULUM

Name of theDegreeProgram : B.Sc. (Basic/Hons.)
DisciplineCore : Biotechnology

Total Credits for the Program : B.Sc. Basic - 136 and B.Sc. Hons. - 176

Starting year of implementation : 2021-22

Program Outcomes:

Competencies need to be acquired by the candidate securing B.Sc. (Basic) or B.Sc. (Hons)

By the end of the program the students will be able to:

Competencies need to be acquired by a candidate securing B.Sc. (Basic) or B.Sc. (Hons) degree in Biotechnology.

- 1. Understanding concepts of Biotechnology and demonstrate interdisciplinary skills acquired in cell biology, genetics, biochemistry, microbiology, and molecular biology.
- 2. Demonstrating the Laboratory skills in cell biology, basic and applied microbiology with an emphasis on technological aspects
- 3. Competent to apply the knowledge and skills gained in the fields of Plant biotechnology, animal biotechnology and microbial technology in pharma, food, agriculture, beverages, herbal and nutraceutical industries.
- 4. Critically analyze the environmental issues and apply the biotechnology knowledge gained for conserving the environment and resolving the problems.
- 5. Demonstrate comprehensive innovations and skills in the fields of biomolecules, cell and organelles, molecular biology, bioprocess engineering and genetic engineering of plants, microbes, and animals with respect to applications for human welfare.
- 6. Apply knowledge and skills of immunology, bioinformatics, computational modelling of proteins, drug design and simulations to test the models and aid in drug discovery.
- 7. Critically analyze, interpret data, and apply tools of bioinformatics and multi omics in various sectors of biotechnology including health and Food.
- 8. Demonstrate communication skills, scientific writing, data collection and interpretation abilities in all the fields of biotechnology.
- 9. Learning and practicing professional skills in handling microbes, animals and plants and demonstrate the ability to identify ethical issues related to recombinant DNA technology, genetic engineering, animals handling, intellectual property rights, biosafety, and biohazards.
- 10. Exploring the biotechnological practices and demonstrating innovative thinking in addressing the current day and future challenges with respect to food, health, and environment.
- 11. Thorough knowledge and application of good laboratory and good manufacturing practices in biotech industries

- 12. Understanding and application of molecular biology techniques and principles in forensic and clinical biotechnology.
- **13.** Demonstrate entrepreneurship abilities, innovative thinking, planning, and setting up small-scale enterprises or CROs.

Continuous Formative Evaluation/Internal Assessment

Total Marks for each course = 100%

Continuous assessment (C1) = 20% marks

Continuous assessment (C2) = 20% marks

Semester End Examination (C3) = 60% marks.

- a) The first component (C1) of assessment is for 20% marks. This shall be based on test, assignment, seminar, case study, field work, project work etc. This assessment and score process should be completed after completing 50% of syllabus of the course/s and within 45 working days of semester program.
- b) The second component (C2) of assessment is for 20% marks. This shall be based on test, assignment, seminar, case study, field work, internship / industrial practicum / project work etc. This assessment and score process should be based on completion of remaining 50 percent of syllabus of the courses of the semester.
- c) During the 17th 19th week of the semester, a semester end examination shall be conducted by the University for each course. This forms the third and final component of assessment (C3) and the maximum marks for the final component will be 60%.
- d) The outline for continuous assessment activities for Component-I (C1) and Component-II (C2) of a course shall be as under.

Outline for continuous assessment activities for C1 and C2

Activities	C1 (% marks)	C2 (% marks)	C1 + C2 (% marks)	C3 (% marks)
Session Test	10	10	20	-
Seminars/Presentations/Assignmen t /Activity	10	10	20	-
Semester end Examination	-	-	-	60
Total	20	20	40	60

• For practical course of full credits, Seminar shall not be compulsory. In its place, marks shall be

awarded for Practical Record Maintenance.(the ratio is 50%: 50%)

- Conduct of Seminar, Case study / Assignment, etc. can be either in C1 or in C2 component at the convenience of the concerned teacher.
- The teachers concerned shall conduct test / seminar / case study, etc. The students should be informed about the modalities well in advance. The evaluated courses/assignments during component I (C1) and component II (C2) of assessment are immediately provided to the candidates after obtaining acknowledgement in the register by the concerned teachers(s) and maintained by the Chairman in the case of a University Post-Graduate Department and the Principal / Director in the case of affiliated institutions. Before commencement of the semester end examination, the evaluated test, assignment etc. of C1 and C2 shall be obtained back to maintain them till the announcement of the results of the examination of the concerned semester.
- e) The marks of the internal assessment shall be published on the notice board of the department / college for information of the students.
- f) The Internal assessment marks shall be communicated to the Registrar (Evaluation) at least 10 days before the commencement of the University examinations and the Registrar (E) shall have access to the records of such periodical assessments.
- g) There shall be no minimum in respect of internal assessment marks.
- h) Internal assessment marks may be recorded separately. A candidate, who has failed or rejected the result, shall retain the internal assessment marks.

Curriculum Structure for the Undergraduate Degree Program

B.Sc. (Basic / Hons.)

Total Credits for the Program : 176 Starting year of implementation : 2021-22

Name of the Degree Program : B.Sc. (Basic/Hons.) BIOTECHNOLOGY

Program Articulation Matrix:

Curriculum Structure for the Undergraduate Degree Program - BSc

Total Credits for the Program: 176

Starting year of implementation: 2021-22

Name of the Degree Program: B.Sc.

Discipline/Subject: Biotechnology

Program Articulation Matrix:

This matrix lists only the core courses. Core courses are essential to earn the degree in that discipline/subject. They include courses such as theory, laboratory, project, internships etc. Elective courses may be listed separately

Pedagogy for student engagement is predominantly lectures. However, other pedagogies enhancing better student engagement to be recommended for each course. The list includes active learning/ course projects/ problem or project-based learning/ case studies/self-study like seminar, term paper or MOOC

\$ Every course needs to include assessment for higher order thinking skills (Applying/ Analyzing/ Evaluating/ Creating). However, this column may contain alternate assessment methods that help formative assessment (i.e. assessment for learning).

BSc Biotechnology (Basic / Hons.) Semester 1

Course Title: DSC-1T, BTC 101, Cell Biology and Genetics					
Total Contact Hours: 56	Course Credits: 4+2				
Formative Assessment Marks:40%	Duration of ESA/Exam: 3 Hrs				
Model Syllabus Authors: Curriculum Committee	Summative Assessment Marks: 60%				

Course Pre-requisite(s): Mention only course titles from the curriculum that are needed to be taken by the students before registering for this course.

Course Outcomes (COs):

At the end of the course the student should be able to:

(Write 3-7 course outcomes. Course outcomes are statements of observable student actions that serve as evidence of knowledge, skills and values acquired in this course)

- 1. Would be able to comprehend the structure of a cell with itsorganelles
- 2. *Can explain the organization of genes and chromosomes, chromosome morphology and itsaberrations

Course Articulation Matrix: Mapping of Course Outcomes (COs) with Program Outcomes (POs 1-12)

Course Outcomes (COs) / Program Outcomes (POs)	1	2	3	4	5	6	7	8	9	10	11	12
1. Would be able to comprehend the structure of a cell with its organelles	*	*			*							
2. Can distinguishbetween the structure of prokaryotic and eukaryotic cell.	*	*			*							
3. Can explain the organization of genes and chromosomes, chromosome morphology and its aberrations	*	*			*							

Course Articulation Matrix relates course outcomes of course with the corresponding program outcomes whose attainment is attempted in this course. Mark 'X' in the intersection cell if a course outcome addresses a particular program outcome.

BSc Biotechnology (Basic / Hons.)

Semester 1

Title of the Courses:

Course 1 : DSC-1T, Cell Biology and Genetics

Course 2: OE 1T, Biotechnology for human welfare

Course 3: SEC 1T, Biotechnological Skills and Analytical Techniques

Course 1 : DSC	-1T, BTC 101,	Course 2:	OE 1T, BTC 301,	Course 3 : SEC 1T, BTC 701,			
Cell Biology a	nd Genetics	Biotechn	ology for human	Biotechnological Skills and			
			welfare	Analytical Techniques			
Number of	Number of	Number	Number of	Number of	Number of lecture		
Theory	lecture	of	lecture	Theory	hours/semester		
Credits	hours/semester	Theory	hours/semester	Credits			
		Credits					
4	56	3	42	1	14		

Content of Course 1: Theory: DSC-1T, BTC 101, Cell Biology and Genetics 56 Hrs

Unit – 1:Cell as a Basic unit of Living Systems and Cellular Organelles 14Hrs

Concept, Development and Scope of Biotechnology. Historical perspectives. Discovery of cell, the cell Theory, Ultra structure of prokaryotic and eukaryotic cell- (Both plant and animal cells),

Surface Architecture: Structural organization and functions of plasma membrane and cell wall of bacteria and plants.

Cellular Organelles: Structure and functions of cell organelles – Endoplasmic reticulum, Golgi complex, Mitochondria, Chloroplast, Ribosomes, Lysosomes, Peroxisomes, Nucleus (Nuclearenvelope with nuclear pore complex, Nucleolus, Nucleoplasm and Chromatin). Vacuole, Cytosol and Cytoskeleton structures (Microtubules, Microfilaments and Intermediate filaments).

Unit- 2. Chromosomes and Cell Division

14Hrs

General Introduction, Discovery, Morphology and structural organization – Centromere, Secondary constriction, Telomere, Chromonema, Euchromatin and Heterochromatin, Chemical composition and Karyotype. Single-stranded and multi- stranded hypothesis, folded- fibre and nucleosome models. Special type of chromosomes: Salivary gland and Lampbrush chromosmes.

Cell Division: Cell cycle, phases of cell cycle, Regulation of cell cycle-, checkpoints and enzymes involved. Significance of cell cycle, interphase nucleus, stages of mitosis and meiosis, achromatic apparatus, synaptonemal complex.Cell Senescence and programmed cell death.

Unit-3.Genetics: 14Hrs

Mendelian Genetics: Introduction and brief history of genetics. Mendelian theory: Laws of inheritance-dominance, segregation, incomplete dominance, codominance with an example. Law of independent assortment, test cross, back cross.

Gene interaction: Deviations to Mendelian inheritance- Supplementary factors: comb pattern in fowls, Complementary genes- Flower colour in sweet peas, Multiple factors—Skin colour in human beings, Epistasis—Plumage colour in poultry (13:3), Multiple allelism: Blood groups in Humans- ABO and Rh.

Maternal Inheritance: Plastid inheritance in Mirabilis, Petite characters in yeast and Kappa particles in Paramecium.

Sex-linked inheritance- Colour blindness, hemophilia, Y-linked traits.

Unit-4.Linkage and Crossing Over

14Hrs

Introduction, Chromosome theory of inheritance, Coupling and repulsion hypothesis, Linkage in maize and Drosophila, Mechanism of crossing over and its importance, chromosome mapping-linkage map in maize.

Chromosomal variations: A general account of structural and numerical aberrations, chromosomal evolution of wheat and cotton.

Mutations: Types of mutations, Spontaneous and induced, Mutagens: Physical and chemical, Mutation at the molecular level, Applications of mutations- plants, animals and microbes.

Sex Determination in Plants and animals: Concept of allosomes and autosomes, XX- XY, XX-XO, ZW-ZZ, ZO-ZZ types.

Human Genetics: Karyotype in man, inherited disorders – Allosomal(Klinefelter syndrome and Turner'ssyndrome), Autosomal (Down syndrome and Cri-Du-Chat Syndrome).

Epigenetics: Plant and humans

Course 1: Practical: DSC-1P, BTC 101, Cell Biology and Genetics

- 1) Study and maintenance of simple and compound microscope
- 2) Use of Micrometer and calibration, measurement of onion epidermal cells and yeast
- 3) Study of stages in mitosis from onion root tips
- 4) Study of stages in meiosis in grasshopper testes/onion or Rhoeo flower buds.
- 5) Mounting of polytene chromosomes
- 6) Buccal smear Barrbodies
- 7) Karyotype analysis Human and Onion Human - Normal and Abnormal - Down and Turner's syndromes
- 8) Isolation and staining of Mitochondria
- 9) Isolation and staining of Chloroplast
- 10) RBC cell count by Haemocytometer
- 11) Simple genetic problems based on theory

Text Books / References

Reference:

- 1. Molecular Biology of Cell Bruce Alberts et al, Garlandpublications.
- 2. Animal Cytology and Evolution- MJD, White Cambridge UniversityPublications
- 3. Molecular Cell Biology-Daniel, Scientific AmericanBooks
- 4. Cell Biology Jack d Bruke, The William TwilkinsCompany
- Principles of Gene Manipulations- Old & Primrose, Black Well Scientific Publications
- 6. Cell Biology-Ambrose & Dorothy M Easty, ELBS Publications
- 7. Fundamentals of Cytology- L. W. Sharp, McGraw HillCompany
- 8. Cytology-Willson&Marrison, ReinformPublications
- 9. Molecular Biology- Christopher Smith, Faber & FaberPublications
- 10. Cell Biology & Molecular Biology EDP De Robertis& EMF Robertis, Saunder College.
- 11. Cell Biology- C.B Powar, HimalayaPublications
- 12. Basic Genetics- Daniel L. Hartl, Jones & Barlett Publishers USA
- 13. Human Genetics and Medicine lark Edward Arnold PLondon
- 14. Genetics Monroe W Strickberger, Macmillain Publishers, New York
- 15. Genes V Benjamin Lewin, Oxford UniversityPress.
- 16. Genes I Benjamin Lewin, Wiley Eastern Ltd., Delhi
- 17. Genes II Benjamin Lewin, Wiley & SonsPublications
- 18. Genes III- Benjamin Lewin, Wiley & SonsPublications
- 19. Principles of Genetics- Sinnott, L.C. Dunn, Dobzhansky, McGraw-Hill.
- 20. Genetics Edgar Altenburg Oxford & IBH publications
- 21. Principles of Genetics E.J. Gardener, M.J. Simmons and D.P. Snustad, John Wiley & SonPublications
- 22. Genetics- P.K.Gupta, Rastogi Publication, Meert, India

Course 2: Theory: OE 1T, Biotechnology for Human Welfare Course 2: OE 1T, Microbial Technology for Human Welfare 42Hrs

Unit – 1:Industry

14Hrs

Introduction, Scope, branches and applications of Biotechnology.

Biotechnology in industry:Industrial production of alcoholic beverage (wine), antibiotic (Penicillin), enzyme (lipase)

Applications of biotechnology in food, detergent and pharmaceutical industries

Unit – 2: Environment

14Hrs

Application of biotechnology in environmental aspects:

Bioremediation: Degradation organic pollutants, hydrocarbons and agricultural wastes, Superbug Bioplastics and Biofuels.

Unit – 3: Forensic and Health Sciences

14Hrs

Application of biotechnology in forensic science:

Solving crimes of murder and rape, paternity testing and theft using DNA finger printing techniques Application of biotechnology in health:

Genetically engineered insulin, recombinant vaccines, gene therapy, diagnostics-ELISA and PCR, human genome project.

References:

- 1. Crueger W and Crueger A. (2000). Biotechnology: A textbook ofIndustrial Microbiology.2nd edition. Panima Publishing Co. NewDelhi.
- 2. Patel AH. (1996). Industrial Microbiology. 1st edition, MacmillanIndia Limited.
- 3. Stanbury PF, Whitaker A and Hall SJ. (2006). Principles ofFermentation Technology. 2nd edition, Elsevier ScienceLtd.
- 4. Environmental Biotechnology, Pradipta KumarMohapatra
- 5. Environmental Biotechnology Concepts and Applications, Hans-Joachim Jordening and Jesef Winter
- 6. B.B. Nanda and R.K. Tiwari, Forensic Science in India: A Vision for the TwentyFirst
 - Century, Select Publishers, New Delhi (2001).
- 7. M.K. Bhasin and S. Nath, Role of Forensic Science in the NewMillennium, University of Delhi, Delhi(2002).
- 8. S.H. James and J.J. Nordby, Forensic Science: An Introduction to Scientific and Investigative Techniques, 2nd Edition, CRC Press, Boca Raton(2005).
- 9. W.G. Eckert and R.K. Wright in Introduction to Forensic Sciences,2nd Edition, W.G.Eckert (ED.), CRC Press, Boca Raton(1997).

Course 3: Theory: SEC 1T, Biotechnological Skills and Analytical Techniques

LEARNING OUTCOMES

- Skill enhancement as per National Occupational Standards (NOS) of Lab Technician/ Assistant Qualification Pack issued by Life Sciences Sector Skill Development Council LFS/Q0509, Level 3.
- Knowledge about major activities of biotech industry, regulations, and compliance, environment, health, and safety (EHS), good laboratory practices (GLP), standard operating procedures (SOP) and GMP as per the industrystandards.
- Demonstrate soft skills, such as decision making, planning, organizing, problem solving, analytical thinking, critical thinking, and documentation.

Course 3: Theory: SEC 1T, Biotechnological Skills and Analytical Techniques 14Hrs

1. Insights into biotechnology industry:

Biotechnology Industry in Indian and Global context - organization in context of large/medium/small enterprises, their structure andbenefits.

2. Industry professional skills to be acquired:

Planning and organising skills, decision-making, problem-solving skills, analytical thinking, critical thinking, team management, risk assessment.

3. Interpersonalskills:

Writing skills, reading skills, oral communication, conflict-resolution techniques, interpretation of research data, trouble shooting in workplace

4. Digitalskills:

Basic Computer Skills (MS Office, Excel, Powerpoint, Internet) for Workplace. Professional Email drafting skills and Powerpoint presentation skills

Analytical Skills inlaboratory:

Solutions: Molarity, Molality, Normality, Mass percent % (w/w), Percent by volume (% v/v), parts per million (ppm), parts per billion (ppb), Dilution of concentrated solutions. Standard solutions, stock solution, solution of acids. Reagent bottlelabel reading and precautions

Course 3: Practicals: SEC 1P, Biotechnological Skills and Analytical Techniques

1. Methods and practices of cleaning and management of lab

Learning and Practice of Integrated clean-in-place (CIP) and sterilize-in-place (SIP) as per industry standards, material requirements for cleaning specific area, equipment, ventilation area, personal protective requirements

2. Procedure of cleaning and storage of Lab ware:

Methodology for storage area, Cleaning procedure and materials to be used for various surfaces. Sign boards, labelling do's& don'ts

Knowledge about standard procedures of cleaning or glass ware, plastic ware. Maintenance of inventory

3. Principles and practices of lab safety:

Knowledge about safety symbols and hazard signs. Personal safety gears, utility, and disposal. Equipment safety protocols, chemical safety protocols. Documentation of chemical and equipment usage records. Handling hazardous chemicals.

4. Best practices of usage and storage of chemicals:

Knowledge and practice in handling of chemicals, labelling and stock maintenance. SOP and material handling. Procedures to maintain chemicals, labelling, storage, and disposal.

5. Record maintenance as per SOPs

Labelling of samples and reagents as per SOPs.

Recording detail of work done for research experiments. Importance of study of manuals, health and safety instructions.

- **6.** Usage and maintenance of basic equipment of biotechnology lab: Principles, calibrations, and SOPs of weighing balances, pH meters, autoclaves, laminar flows and biosafety cabinets (levels), basic microscopes, homogenizers, stirrers, colorimeters, UV, and Visible spectrophotometers.
- 7. **Preparation of solutions and standards -** Properties and uses of chemicals commonly used in life sciences laboratories. Maintaining safety standards for handling various solutions and chemicals. Preparation of test reagents and buffers, Protocols for proper mixing of chemicals. Safety precautions while preparation and storage of incompatible chemicals and reagents.
- **8. Preparation of media**: Maintenance and storage of purified water for media (Plant Tissue culture media, Microbiological media, and Animal cell culture media) preparation. Preparation and storage of concentrated stock solutions. Documentation and disposal of expired stocks.

Collection of indents of media requirement, preparation, and storage. Media coding, documentation, and purpose of usage.

9. Practical methods for decontamination and disposal:

Decontamination methods, Safe disposal practices of decontaminated media or materials.

10. Laboratory record writing

Method of record writing , data collection and recording , reporting of result, discussion of result , summary writing, effective power point presentation taking any experiment as example

11. Industry visit or Analytical laboratory visit

Pedagogy:

The general pedagogy to be followed for theory and practicals are as under. Lecturing, Tutorials, Group/Individual, Discussions, Seminars, Assignments, Counseling, Remedial Coaching. Field/Institution/Industrial visits, Hands on training, Case observations, Models/charts preparations, Problem solving mechanism, Demonstrations, Project presentations, Experiential documentation, and Innovative methods.

Active learning as per LSSSDC (NSDC) LFS/Q0509 guidelines, at skill training Level

3. Case studies about application of microbial biomolecules in various industries. Seminar on topics of microbial biochemistry

BSc Biotechnology (Basic / Hons.) Semester 2

Title of the Courses:

Course 1: DSC-2T, Microbiological Methods and Techniques

Course 2: OE-2T, Applications of Biotechnology in Agriculture

Course 1: DSC-2	T, BTC 102,	Course 2: OE- 2T, BTC 302,				
Microbiologica	Methods and	Applications of Biotechnology in				
Techniques		Agriculture				
Number of Theory	Number of lecture	Number of Theory	Number of lecture			
Credits	hours/semester	Credits	hours/semester			
4	56	3	42			

Content of Course: DSC-2T, BTC 102, Microbiological Methods and Techniques 56 Hrs

Unit - 1 General Microbiology and Instrumentation

14Hrs

General Introduction to Microbiology: Scope and relevance of microbiology, important contributions by Robert Koch, Leeuwenhoek, Jenner, Pasteur, Flemming, Ivanowsky

General account on structure, classification and reproduction of bacteria, virus and fungi

Microscopy: Principles and applications of Compound microscope, Dark field microscope, Phase contrast microscope, Fluorescence Microscope, Confocal microscope, Electron Microscopes- TEM and SEM.

Analytical techniques: Working principles and applications: Centrifuge, Ultracentrifuge, Spectrophotometer, Chromatography: Paper, TLC, Column (adsorption, gel-filtration, ion exchange, affinity), HPLC, GC.

Unit - 2 Sterilization techniques

14Hrs

Definition of terms-sterilization, disinfectant, antiseptic, sanitizer, germicide, microbicidal agents, microbiostatic agent and antimicrobial agent.

Physical methods of control: Principle, construction and applications of moist heat sterilization-Pasteurization, Boiling, Fractional sterilization-Tyndallization and autoclave. Dry heat sterilization-Incineration and hot air oven.

Filtration – Diatomaceous earth filter, seitz filter, membrane filter and HEPA

Radiation: Ionizing radiation-y rays and non ionizing radiation- UVrays

Chemical methods: Alcohol, aldehydes, phenols, halogen, metallic salts, Quaternary ammonium compounds and sterilizing gases as antimicrobial agents.

Unit – 3: Microbiological techniques

14Hrs

Culture Media: Components of media, natural and synthetic media, chemically defined media, complex media, selective, differential, indicator, enriched and enrichmentmedia

Pure culture methods: Serial dilution and plating methods (pour, spread, streak); cultivation, maintenance and preservation/stocking of pure cultures; cultivation of anaerobicbacteria

Microbial growth and its measurements: Growth curve, enumeration methods (turbidity, cell counting, colony counting)

Stains and staining techniques: Principles of staining, Types of stains-simple stains, structural stains, negative stain and differential stains.

Unit – 4: Antimicrobial agents

14Hrs

Antibiotic sensitivity testing methods: Disc and Agar well diffusion techniques

Five modes of action with one example each: Inhibitor of nucleic acid synthesis; Inhibitor of cell wall synthesis; Inhibitor of cell membrane function; Inhibitor of protein synthesis; Inhibitor of metabolism

Antifungal agents: Mechanism of action of Amphotericin B, Griseofulvin

Antiviral agents: Mechanism of action of Amantadine, Acyclovir, Azidothymidine Antibiotic resistance, MDR, XDR, MRSA, NDM-1

Course 1: Practicals: DSC-2P, Microbiological Methods and Techniques

- 1. To study the principle and applications of important instruments (biological safety cabinets, autoclave, incubator, BOD incubator, hot air oven, light microscope, pH meter) used in the microbiology and Biotechnologylaboratory.
- 2. Sterilization of medium using Autoclave and assessment forsterility
- 3. Sterilization of glassware using Hot Air Oven and assessment forsterility
- **4.** Sterilization of heat sensitive material by membrane filtration and assessment for sterility
- **5.** Preparation of culture media for bacteria, fungi and theircultivation.
- **6.** Plating techniques: Spread plate, pour plate and streak plate.
- 7. Isolation of bacteria and fungi from soil, water andair
- 8. Study of Rhizopus, Penicillium, Aspergillus using temporarymounts
- 9. Colony characteristics study of bacteria from air exposureplate
- **10.** Staining techniques: Bacteria– Gram, Negative, Capsule, Endospore staining Fungi Lactophenol cotton bluestaining
- 11. Water analysis MPNtest
- 12. Biochemical Tests IMViC, Starch hydrolysis, Catalase test, Gelatinhydrolysis
- 13. Bacterial cell motility hanging drop technique

Text Books / References

- 1. Atlas RM. (1997). Principles of Microbiology. 2nd edition. WM.T. Brown Publishers.
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- 15. Frontiers in Microbial technology-P.S. Bison, CBSPublishers.
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Course 2: Theory: OE- Applications of Biotechnology in Agriculture 42 Hrs

Unit – 1: Agricultural Biotechnology

14 Hrs

Concepts and scope of biotechnology in Agriculture. Plant tissue culture, micro propagation, entrepreneurship in commercial plant tissue culture. Banana tissue culture - primary and secondary commercial setups, Small scale bioenterprises: Mushroom cultivation

Unit − **2**: Transgenic plants

14 Hrs

The GM crop debate – safety, ethics, perception and acceptance of GM crops

GM crops case study: Bt cotton, Bt brinjal, Biopesticides: Baculovirus pesticides, Mycopesticides Genetic Engineering for quality improvement: Golden rice, Seed storage proteins, Flavours—capsaicin, vanillin

Unit – 3: Molecular pharming and post harvest protection

14 Hrs

Plants as biofactories for molecular pharming: edible vaccines, plantibodies, nutraceuticals Post-harvest Protection:

Antisense RNA technology for extending shelf life of fruits and shelf life of flowers. Biosafety, bioethics and IPR.

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GENERAL PATTERN OF THEORY EXAMINATION

B.Sc – BIOTECHNOLOGY

Theory Question Paper Pattern for DSC, DSE, and OE Courses

Duration: 3 Hours Maximum: 60 Marks All questions are compulsory Draw neat labeled diagrams wherever necessary QNo. I Answer any EIGHT of the following 2X8=16 (1) (2) (3) (4) (5) (6) (7) (8) (9) (10)QNo. II Answer any SIX of the following 4X6=24(11)(12)(13)(14)(15)(16)(17)(18)QNo. III Answer any TWO of the following 10X2=20(19)(20)(21) (22)

PATTERN OF PRACTICAL EXAMINATION

Practical examination – B. Sc BIOTECHNOLOGY

Duration: 3 hours	Max. Marks: 25

Q. 1 Major question 08 Marks

Q. 2 Minor question 05 Marks

Q. 3 Identify and comment 2X4 = 08Marks

Q. 4 Viva-voce 04 Marks

PATTERN OF FORMATIVE

ASSESMENT – PRACTICALS Max. Marks: 25

1 IA 1 10 Marks

2 IA 2 10 Marks

3 Record 05 Marks
